

# The effects of tissue plasminogen activators on experimental cerebral ischemic infarcts

Kayhan KUZEYLİ<sup>1</sup>, Savaş CEYLAN<sup>1</sup>, Süleyman BAYKAL<sup>1</sup>,  
Konuralp İLBAY<sup>1</sup>, Ertuğrul ÇAKIR<sup>1</sup>, Soner DURU<sup>1</sup>, Müfit KALELİOĞLU<sup>2</sup>,  
Fadıl AKTÜRK<sup>1</sup>, Sezer Ş.KOMSUOĞLU<sup>3</sup>, Yavuz ÖZORAN<sup>4</sup>

<sup>1</sup>Göztepe SSK Hospital, İstanbul, Depts. of neurosurgery, <sup>2</sup>Neurology and <sup>3</sup>Pathology, Medical School of KTU, Trabzon, TURKEY

*In this study, the effects of tissue plasminogen activators (TPA) on experimental cerebral embolic ischemia and/or infarcts in rats were investigated. By administering TPA IV bolus fortyfive minutes after embolus material that will cause experimental cerebral embolic ischemia and/or infarcts had been given into the internal carotid artery; a significant improvement in neurological deficits was observed ( $p < 0.05$ ). In the histopathological evaluation; it was observed that TPA caused a significant decrease in the size of ischemia and/or infarct areas ( $p < 0.05$ ) but did not cause any bleeding in them. In this study it was also concluded that, application of TPA as intra-venously bolus "which is different from the classical way" is useful in embolic strokes and does not cause a significant complication. [Turk J Med Res 1994; 12(1): 5-10]*

Key Words: Embolism, Infarct, Ischemia, Tissue Plasminogen Activator (TPA), Rat

The most important part of the cerebral ischemic infarcts are results of thromboembolies (1,2).

If regional ischemia which is due to the regional cut off of the cerebral blood flow (cbf) by thromboembolies is not abolished immediately, that will result in irreversible tissue necrosis and cerebral infarct. It is known that, in recent years the importance of fibrinolytic (thrombolytic) treatment methods are increasing (3,17). If there is thrombus, TPA turns, inactive plasminogen to active plasmin by provoking the fibrinolytic enzymes. This property is specific to clot (5,7,11). The lysis of the clot occurs by this way.

TPA's selectivity to clot inhibits systemic activation of the fibrinolytic system and this causes lesser damage in hemostatic system. In addition to that when it is compared to the other fibrinolytic agents its complications are rare (3,15,18,31).

## MATERIALS AND METHODS

In the study, 26 Wistar rats weighing 200-300 g were used. The rats didn't take anything by mouth for 6 hours before operation and their body temperatures were kept. For anesthesia intraperitoneal ketamin and

0.1 ml IM atropine were used. After the anesthesia, the manipulation was begun while the rat was respirating spontaneously and fixated at the supine position. The muscles were passed by skin incision which was made just left of the midline between the xyphoid and crichoid cartilages. Under the operation microscope the carotis sheath was dissected ten minutes after the local anesthetic was dropped on it. Then, the internal and external carotid arteries were exposed. Later left yoid bone was extracted partially and the pterygopalatal branches of external carotid artery were electrocauterised. Embolus material was prepared by the modification of the technic suggested by Kaneko et al (9). Blood which was taken 24 hours before from a healthy rat by cardiac puncture was kept at 37 °C. 0.25-0.50 mm<sup>3</sup> of it was mixed with 0.3 ml saline and injected slowly and caudally into the external carotid artery which was held up at the distal end through the common carotid bifurcation with 27 G catheter, while the catheter was taken out; external carotid artery held up from the suture proximal of it and the hole of the catheter was cauterised.

After internal carotid artery was seen intact, one layer closure was made.

In the study group, 45 minutes after the embolus material had been injected, 200 IU TPA/ml was given in 5 minutes from the canulised femoral vein (32). Rats in both were observed for ten minutes after the operation for their vital signs in the recovery cage in the semifowler position. All rats were evaluated in 6

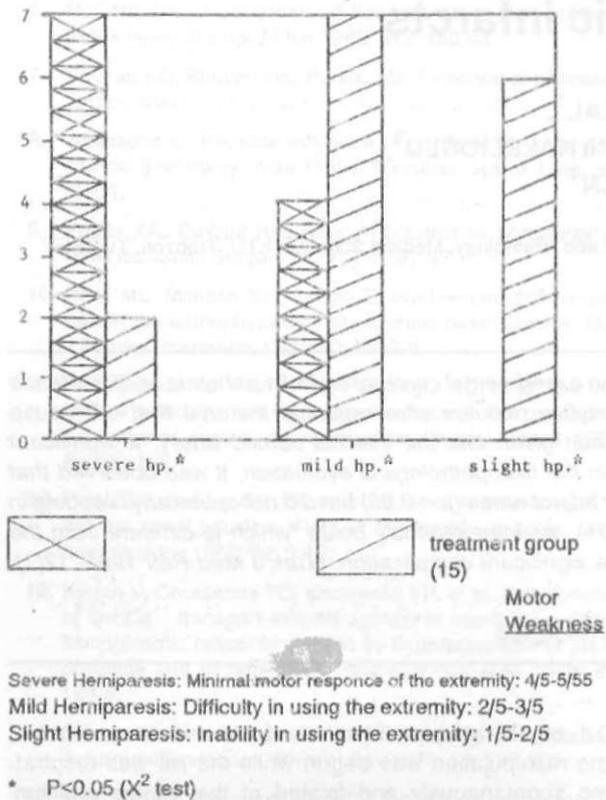
Received: July, 13, 1993

Accepted: Dec. 28, 1993

Correspondence: Kayhan KUZEYLİ

Dept. of Neurosurgery, Medical School of  
KTU, Trabzon, TURKEY

table 1. Neurologic grading of the rats at the control and treatment groups.



hours intervals with the modifications of the neurological scales used by Zvin et al, Threepoint (16) and Pang group. Rats which had neurological deficits at the end of the 24 hours were included to the study and normal ones were excluded. The rats in the study group were sacrificed 24 hours later under ether anesthesia. Their brains were extracted by craniectomy. The extracted brains were fixated in %10 phosphate tamponade solution for 15 days.

After the fixation procedures, the brains were sliced in coronal plane from front to back (middle frontal-parietal-cerebellar) and the tissue samples were stained with hematoxylin-eosin (HE) for histopathological evaluation after usual processing.

## RESULTS

There were ill rats in the control group. 7 (%64) had severe, 4 (%36) had mild hemiparesis. There were 15 rats in the study group. TPA was administered to these rats, 2 (%14) of them had severe, 7 (%46) of them had mild, 6 (%40) of them had slight hemiparesis (Table I).

The rats in the control group had a mean PTT value of 28.63 sec. In the control mean PT value was 23.89 sec. and the mean PTT VALUE WAS 28.63

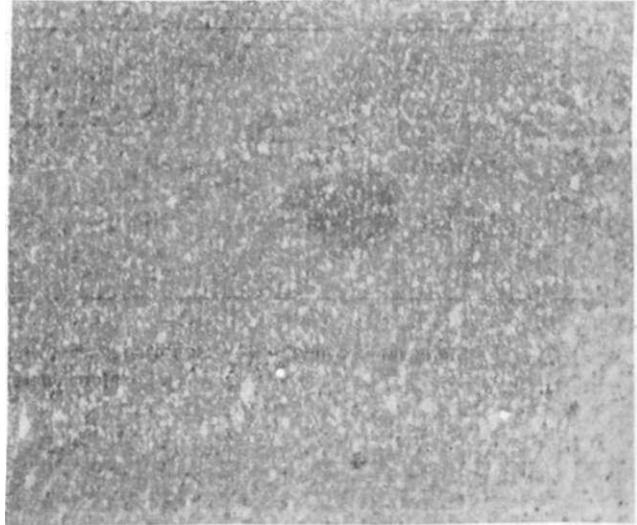


Figure 1. Supratentorial brain sections of the control group. Ischemic infarct area occurred by softening in the white matter, pale neurophilic, edema and necrosis (HE x 12.5).

sec. In the histopathological examination of the rat (brains) in the control group; ischemic infarct zones which were emerging from softening of the supratentorial cerebral parenchyma, pale neurophilic; edema and necrotic remnants had been observed (Figure 1). In ipsilateral supratentorial regions 6 rats had 3, 5 rats had 2 different infarct zones (Figure 2). However, in

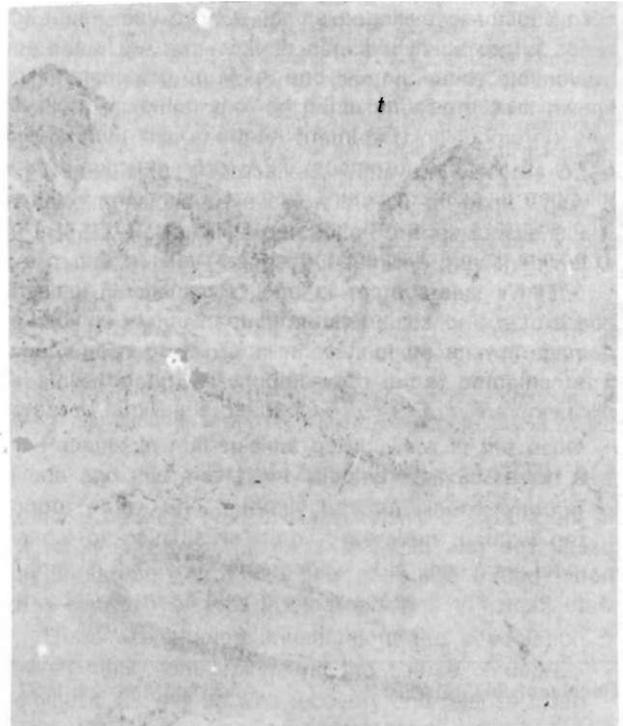


Figure 2. Multiple ischemic infarct areas at the supratentorial brain sections of the control group (HE x 3.2).

THE EFFECTS OF TISSUE PLASMINOGEN ACTIVATORS ON EXPERIMENTAL CEREBRAL ISCHEMIC INFARCTS

Table 2 Micrometric sizes of the cerebral infarcts at the control and treatment groups\*

	Unit scale	Micrometric Scale	
1	35.35	1319.	1319
	20.23	1055,6.	867.
	10.18	377.	378,6.
2	13.22	490.	029,4.
	10.20	377.	754.
	5.15	188,5.	565,5.
3	12,27	452,4.	1017,5.
	5,9	188,5	339.
	14,1 i	527,8	414,7.
4	10,20	377.	1055,6
	15,23	565,5.	867.
	18,20	678,6	754.
5	20,15	754.	565,5.
	13,15	490	565,5.
	6,10	226.	377.
6~	23,20	867	754.
	14,22	527,8.	829,4.
7	28,30	1055,6.	1131.
	20,15	754.	565,5.
tT	20,15	754.	565,5.
	j8,10	678,6.	377.
10	12,22	452,4.	829,4.
		678,6.	754.
11	20,22	754.	629.
	17,26	640.	980.
11	13,15	490.	565,5
	10,15	377.	565,5.
	18,8	670,6.	301,6.

\* One distance is equal to 37.7 Micron (x10,4)

the treatment group 10 rats had 3, 5 rats had 2 different ipsilateral supratentorial infarcts (Table II). The infarct zones which were observed in the study group had widespread inflammatory cell infiltration beside the infarct signs in the control group (Figure 3,4). The dimension of the ischemic infarcts which were examined micrometrically in the control group were; the biggest 1319;1319 micron, the least 188,5;113 micron (Table II, III). Neither microscopically nor macroscopically hemorrhage haven't been observed in both groups.

**DISCUSSION**

There are two handicaps in the evaluation and classification of the experimental cerebral embolies.

First is the determination of the observation period after the embolic material had been given. Thus, in many cerebral infarcts which were obtained by clipping or ligating the main cerebral arteries or injecting the embolus material directly into the cerebral artery. It was observed or declared that neurological deficits could recover or improve within a few days spontaneously. It is known that this is due to the

Treatment Groups

	Unit Scale	Micrometric Scale	
1	5,5	188,5.	188,5
	3,5	11,3	188,5.
	8,14	301,6	527,8.
2	10,5	377.	188,5.
	3,5	113.	138,5.
	3,3	113.	188,5
3	15,12	565,5	152,4.
	3,0	301,6.	377.
4	8,8	301,6.	377.
	10,12	377.	452,4.
	8,11	301,6	414,7.
5	11,12	414,7.	452,4.
	4,5	160,8.	188,5
	5,5	188,5.	183,5.
6	17,17 "	301,6.	640,9.
	3,3	113.	113.
	5,4	188,5.	150,8.
7	3,4	113.	150,8.
	3,3	113.	113.
	6,10	301,6	377.
8	13,3	490.	400.
	3,4	113.	150,8.
9	8,8	301,6.	301,6.
	5,6	188,5	226.
	4,8	150,8.	301,6.
10	10,10	377	377.
	5,3	188,5.	113.
	7,10	263,2.	377.
11	18,12	678,6.	452,4.
	6,6	226.	226.
12	5,9	188,5.	389.
	9,11	339.	414,7.
	3,5	113.	188,5.
13	8,15	301,6	565,5.
	3,4	113.	150,3.
	10,6	377.	226.
14	11,11	414,7.	414,7.
	8,8	301,6.	301,6.
15	8,8	301,6.	301,6.
	4,9	150,8.	339.

decrement of the ischemic brain edema at postoperative 3-4 days or contralateral hemisphere undertakes the functions of the ipsilateral hemisphere (33-34).

The second handicap is the timing of the incubation period of the embolus material. This period is important, because this can lead us to false results in fibrinolytic treatment (13).

In the light of these two factors; In our study rats were sacrificed 24h after embolus material had been given. In addition to that by incubating embolus material for 24 hours, the fibrinolytic activity in the fresh clot was tried to be minimized.

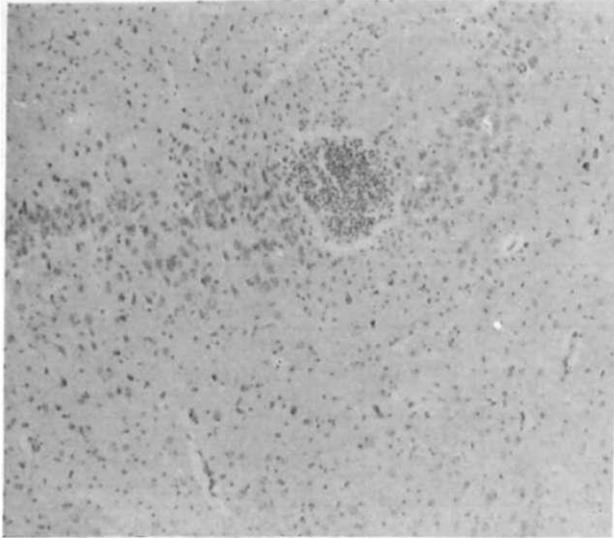


Figure 3. Supratentorial brain sections of the treatment group. Significant inflammatory cell infiltration at the infarct areas (HE x 12.5).

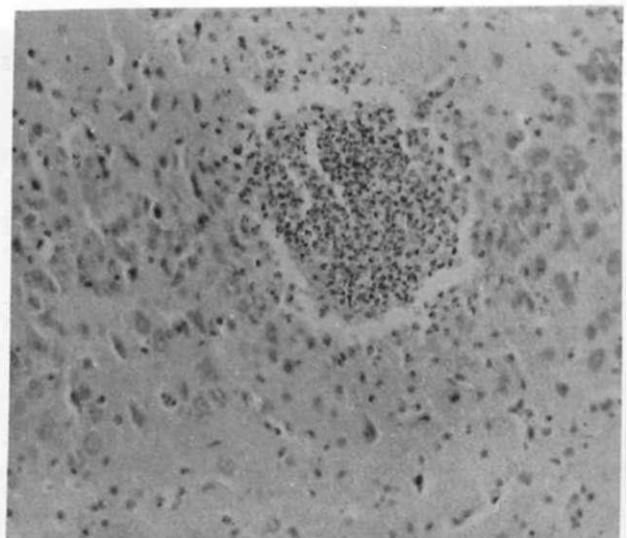
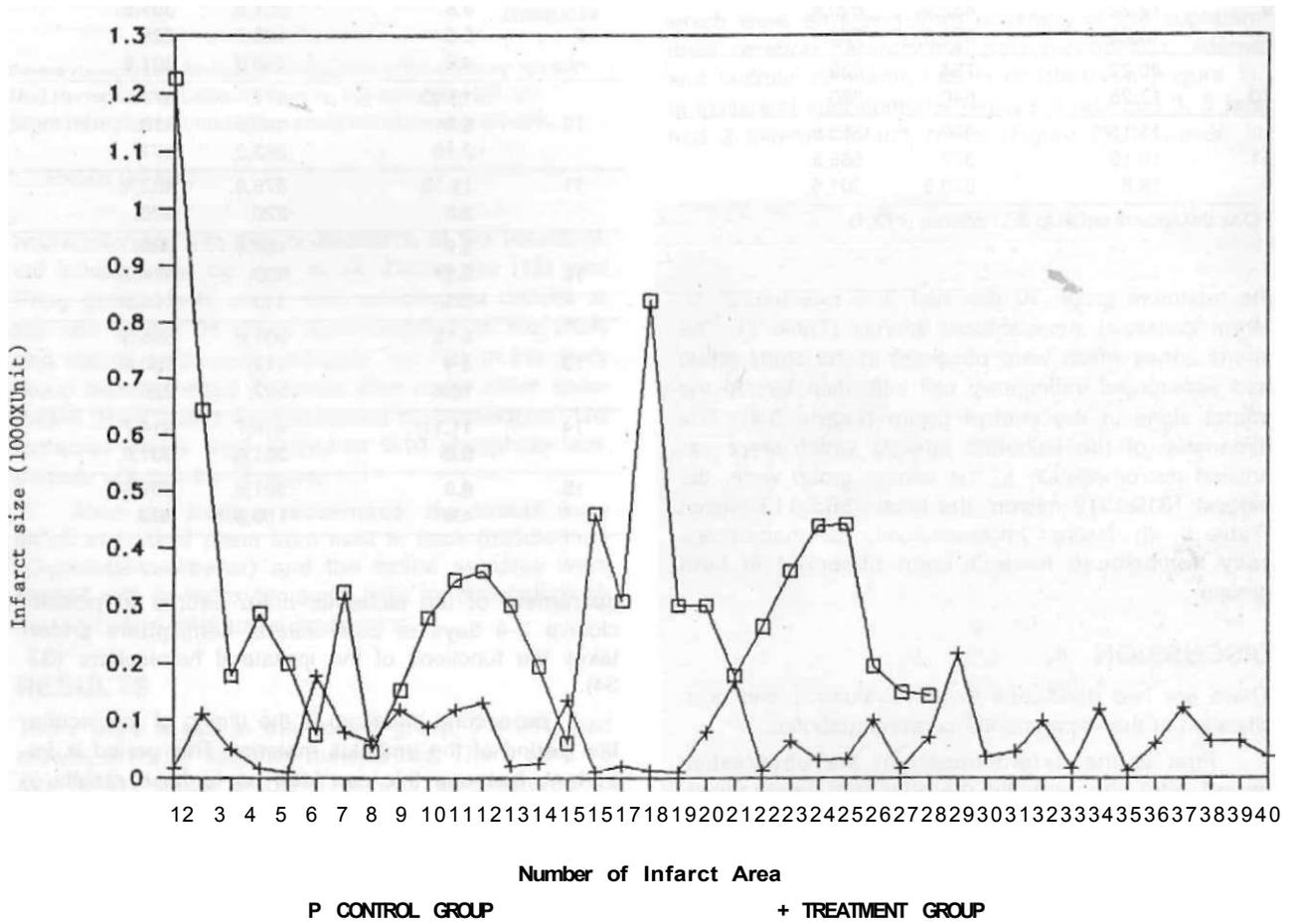


Figure 4. Magnified view of the ischemic infarct zones (HE x 25).

Table 3. Comparison of the infarct sizes.



In the fibrinolytic treatment total dose of TPA was administered gradually. Sixty percent of total dose was given as quick intravenous infusion to obtain therapeutic dose that establishes reperfusion and remaining %40 was infused with 30-90 minutes intervals as continuous perfusion to decrease the reocclusion incidence. Although the perfusion treatment is suggested. It is also known that it affects the coagulation system negatively (38).

In our study by giving TPA's total dose IV in 5 minutes, we tried to overcome coagulation defect and have maximal lytic effect of the fibrinolytic agent.

In the presence of fibrin TPA turns plasminogen to plasmin and provides an effective thrombolysis. By this way it opens %60 of occluded cerebral vessel and increases the regional ischemic flow (39).

Papadopoulos (40), Kissel (36), Zivin (16) and other researchers showed in their experimental cerebrovascular studies that TPA increases the blood flow in embolic vessels (23).

When both groups in our study were compared, we observed that improvement in the neurological deficits of the rats in the treatment group were statistically significant ( $p<0.05$ ) (Table I). When both groups in our study were compared, we observed that improvement in the neurological deficits of the rats in the treatment group were statistically significant ( $p<0.05$ ). This histopathological improvement was parallel to the clinical improvement. It is possible that TPA can cause these effects by lysing the intravascular embolus and restoring the blood flow in the ischemic zone (36,39,40). Therefore, as idling neurons in penumembrana gain function, improvement both histopathologically and clinically ensues.

On the other hand none of the infarcts in the study group neither showed hemorrhage nor turned to hemorrhagic infarcts. It is known that, although continuous infusion of TPA minimizes reocclusion risk, it affects the coagulation system negatively (38). In our study by giving TPA rapidly in six hours we provided a maximal lytic effect on the embolus but didn't have a negative effect on the coagulation system and this correlates well with the reports saying that, TPA does not provoke changing of ischemic infarcts to hemorrhagic infarcts (13,24,28,41,42). By giving TPA as bolus, TPA did not change the nature of the ischemic infarcts in the study group.

According to these data, we can say that; under the general indications of fibrinolytic treatment, giving the total TPA dosage IV in a short time will improve the neurological deficits in the cerebral ischemic infarcts and won't cause any important complications.

*Turk J Med Res 1994; 12(1)*

### **Doku plazminojen aktivatörlerinin deneysel serebral infarktlar üzerine etkisi**

*Bu çalışmada, doku plazminojen aktivatörlerinin (TPA) ratlarda oluşturulan deneysel serebral embolik iskemi ve/veya infarktlara etkisi araştırıldı. Internal karotisiarterine araştırıldı. Embolus materyali verildikten 45 dk. sonra intravenöz bolus tarzında TPA verilerek nörolojik desifitlerde anlamlı bir iyileşme belirlendi ( $P<0.05$ ). Histopatolojik değerlendirmede TPA'nın iskemi ve/veya infarkt alanlarında anlamlı derecede azalmaya (ancak kanama yapmadığı) yol açtığı görüldü ( $p<0.05$ ).*

*Bu çalışmada TPA'nın klasik yoldan farklı olarak intravenöz bolus tarzında uygulamasının embolik inmelerde yararlı olacağı ve önemli bir komplikasyona yol açmayacağı soucuna ulaşılmıştır. [TurkJMedRes 1994; 12(1): 5-10]*

### **REFERENCES**

1. Chambers BR, Donnan GA, Bladen PF. An analysis of the first 700 consecutive admissions to the Austin Hospital Stroke Unit. *Nz J Med* 1983; 13:57-74.
2. Herman B, Leyten ACM, Lujik JH, et al. Epidemiology of stroke in Tilburg. The Netherlands. *Stroke*. 1982; 13:629-34.
3. Agnelli G, Buchanan MR, Fernandez F et al. A comparison of the thrombolytic and hemorrhagic effects of tissue-type-plasminogen activator and streptokinaz in rabbits. *Circulation* 1985; 72-1:178-82.
4. Collen D, Lijnen HR. New approaches to thrombolytic therapy. *Arteriosclerosis* 1984; 4:579-85.
5. Collen D, Lijnen HR. Tissue type plasminogen activator mechanism of action and thrombolytic properties. *Haemostasis* 1986; 16:25-32.
6. Garabedian HD, Gold HK, Leinbach RC et al. Dose dependent thrombolysis, pharmacokinetics and hemostatic effects of recombinant human tissue-type plasminogen activator for coronary thrombosis. *Am J Cardiol* 1986; 58:673
7. Haylaerts M, Rijken DC, Lijnen HR, et al. Kinetics of activation plasminogen by human tissue plasminogen activator. Role of fibrin. *J Biol Chem* 1982; 257:2912-9.
8. Jones TH, Morawetz RB, Crowell RM, et al. Threshold of focal cerebral ischemia in awake monkeys. *J Neurosurg.* 1981;54:773-82.
9. Kaneko D, Nakamura N, Ogawa T. Cerebral infarction in rats, using homologous blood emboli, development of a new experimental model. *Stroke* 1985; 16(1):76-84.
10. Matsuo O, Rijken DC, Collen D. Thrombolysis by human tissue plasminogen activator and urokinase in rabbits with experimental pulmonary embolus. *Nature* 1981; 291:590-1.
11. Rijken DC, Haylaerts M, Collen D. Fibrinolytic properties of one-chain human extrinsic (tissue-type) plasminogen activator. *J Biol Chem* 1982; 257:2920-5.
12. Seki H, Yohimoto T, Ogawa A, et al. The CO<sub>2</sub> response in focal cerebral ischemia, sequential changes following recirculation. *Stroke* 1984; 15:699-704.

13. Sobel BE, Gross RW, Robinson AK. Thrombolysis clot selectivity and kinetics. *Circulation* 1984;70:160-4.
14. Todd NV, Picozzi P, Crockard HA, et al. Reperfusion after cerebral ischemia influence of duration of ischemia. *Stroke* 190C: 17: 460-6.
15. Zeumer H. Vascular recanalizing techniques in intervention? *J Neurology* 1985; 231: 287-94.
16. Zivin JA, Lyden PD, De Giromali U, et al. Tissue plasminogen activator. Reduction of neurologic damage after experimental embolic stroke. *Arch Neurol* 1988; 45: 337-91.
17. Zivin JA, Fisher M, De Girolami U, et al. Tissue plasminogen activator reduces neurological damage after cerebral embolism. *Science* 1985; 230:1289-92.
- 10 Bergmann SR, Pox KA, et al.. Clot-selective coronary thrombolysis with tissue plasminogen activator. *Science* 1983;330:1181-3.
19. Gaffe GL, Brannward E. Thrombolytic therapy. A new strategy for the treatment of acute myocardial infarction. *N Eng J Med* 1984; 311: 770-6.
20. De Zappo GJ, Copeland BR, Walts TA, et al. The effect of intracerebral urokinase on acute stroke in a baboon model. *Stroke* 1986; 17: 63: \*3.
21. De Zappo GJ, Zeumer M, Harkness LA. Thrombolytic therapy in stroke; possibilities and hazards. *Stroke* 1986 Jul-Aug; 17:595-607.
22. Hinnaway J, Torack R, Fletcher AP, et al. Intracranial bleeding associated with urokinase therapy for acute ischemic stroke. *Stroke* 1976; 7:143-G.
23. Lytton DP, Zivin JA, Clark WA, et al.. Tissue plasminogen activator mediated thrombolysis of cerebral emboli and its effect on hemorrhagic infarction in rabbits. *Neurology* 1979; 29: 703-8.
24. Matsuo O, Rijken P, Colten D. Comparison of the relative fibrinolytic and thrombolytic properties of tissue plasminogen activator and urokinase in vitro. *Thromb Haemostasis* 1981; 45: 225-9.
25. Matsuo O, Kosigi T, Mihara H, et al. Retrospective study on the efficacy of using urokinase therapy. *Nippon Ketsumu Gakkaishi* 1979; 47: 684-8.
26. Philips DA, Fischer M, Smith 7W, et al. The safety and angiographic efficacy of t-PA in a cerebral embolism model. *Ann Neurol* 1988; 23: 391-4.
27. The TIMI study group. The thrombolysis in myocardial infarction\* (TIMI) trial. Phase-I. Findings. *N Eng J Med* 1985; 312:9-12 6.
28. Vande Werf F, Ludbrook PA, Bergmann SR. Coronary thrombolysis with tissue type-plasminogen activator in patients with evolving myocardial infarction. *N Eng J Med* 1904; 310:609-13.
29. Vorstaele M, Bory M, Collen P. Randomized trial of intravenous recombinant tissue type plasminogen activator versus intravenous streptokinase in acute myocardial infarction. *Lancet* 1985. 1:042-7.
30. Weimer W, Stibbe J, Van Seyen A, et al. Specific lysis of an iliofemoral thrombus by administration of extrinsic (tissue-type) plasminogen activator. *Lancet* 1981; 11:1018-20.
31. Weinfeld FD. National survey of stroke. *Stroke* 1981; 12(1): 1-15.
32. Kaufman JJ, Schochet S, Koos V, et al. Efficacy and safety of tissue plasminogen activator. *Neurosurgery* 1987; 20: 403-7
33. Little JR, Sundz TM Jr, Kerr FWL. Neuronal alterations in developing cortical infarction. An experimental study in monkeys. *Neurosurgery* 1974; 40: 180-98.
34. Redding RW. Anatomy and physiology in Hoedlein BP. *Canine Neurology Diagnosis and Treatment* ed. 3. Philadelphia. WB Saunders 1970; 7-51.
35. Tiefenbrun At, Sobel BE. t-PA activator- an agent with promise for selective thrombolysis. *Int J Cardiol* 1985; 7: 32-6.
36. Kissel P, Chehrizi B, Seibert JA. et al. Digital angiographic quantification of blood flow dynamics in embolic stroke treated with tissue-type plasminogen activator. *J Neurosurg* 1987; 67:399-406.
37. Mueller HS, Roberits P, Teichman SI., et al. Thrombolytic therapy in acute myocardial infarction Part: II. medical Clinics of North America 1989 March; 73(2)- 387-407.
38. Gold NK, Loinbach RC, Carabedian HD. et al. Acute coronary reocclusion after thrombolysis with recombinant human tissue plasminogen activator. *Circulation* 1986; 72: 347-52.
39. Chehrizi B, Seibert JA, Kissel P, et al. Evaluation of recombinant tissue plasminogen activator in embolic stroke. *Neurosurgery* 1989; 24: 355-60.
40. Papadopoulos UM, Chendler WF, Topal EF, et al. Recombinant human tissue plasminogen activator therapy in acute thromboembolic stroke. *J Neurosurg* 1987; 67: 394-8.
41. Carlson SE, Aldrich SM, Greenberg HS, et al. Intracerebral hemorrhage complicating intravenous tissue plasminogen activator treatment. *Arch Neurol* 1988; 45: 1070-3.
42. Castellino FJ. Biochemistry of human plasminogen. *Semin Thromb Hemostasis* 1984; 10:18-23.